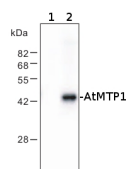


Product no **AS09 485****MTP1 | vacuolar Zn²⁺/H⁺ antiporter****Product information****Immunogen** | KLH-conjugated synthetic peptide derived from *Arabidopsis thaliana* MTP1 UniProt:Q9ZT63, TAIR:At2g46800**Host** | Rabbit**Clonality** | Polyclonal**Purity** | Serum**Format** | Lyophilized**Quantity** | 100 µl**Reconstitution** | For reconstitution add 100 µl of sterile water**Storage** | Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.**Additional information** | 0.1 % sodium azide is added as preservative. For antibody re-suspending information check the tube label.

MTP1 protein is of low abundance in plant tissues.

Application information**Recommended dilution** | 1 : 8000 (ELISA), 1 : 1000 (WB)**Expected | apparent MW** | 43.8 | 43 kDa (*Arabidopsis thaliana*)**Confirmed reactivity** | *Arabidopsis thaliana*, *Nicotiana tabacum***Predicted reactivity** | *Brassica sp.*, *Noccaea fendleri*, *Thlaspi caerulescens*
Species of your interest not listed? [Contact us](#)**Not reactive in** | *Hordeum vulgare*, *Solanaceae sp.***Additional information** | Protein or membrane sample should be treated at 70°C for 10 min before loading on the gel**Selected references** | [Vera-Estrella et al. \(2017\)](#). Cadmium and zinc activate adaptive mechanisms in *Nicotiana tabacum* similar to those observed in metal tolerant plants. *Planta*. 2017 Apr 28. doi: 10.1007/s00425-017-2700-1.
[Kawachi et al. \(2008\)](#). Deletion of a histidine-rich loop of AtMTP1, a vacuolar Zn(2+)/H(+) antiporter of *Arabidopsis thaliana*, stimulates the transport activity. *J.Biol. Chem.* 13:8374-8383.
[Kobae et al. \(2004\)](#). Zinc transporter of *Arabidopsis thaliana* AtMTP1 is localized to vacuolar membranes and implicated in zinc homeostasis. *Plant Cell Physiol.* 12:1749-1758.**Application example**

Sample of a vector (1) and vacuolar membrane fraction of yeast cells expressing AtMTP1 (2) separated on 12 % SDS-PAGE and blotted 1h to PVDF membrane (40 min. at 10 V using BioRad semidry transfer). Filters were blocked 1h with 5 % low-fat milk powder in TBS-T (0.05% Triton X.100). Membranes were washed 5 times with TBS-T, each time in a fresh polystyrene box and probed with anti-AtMTP1 antibodies (AS09 485, 1:1000, 1h) and secondary anti-rabbit (1:2000, 1 h). All steps were performed in RT with agitation.