

This product is for research use only (not for diagnostic or therapeutic use)

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Product no AS15 2898 Anti-Ferritin 1-2 (plant) Product information

Immunogen	Purified ferritin from dried peas, <i>Pisum sativum</i> L. After extraction from pea flour, the ferritin was further purified by gel filtration chromatography to >95% purity as estimated from a Coomassie-stained gel. Antibody is most likely to bind to all ferritin isoforms from pea however it has not been confirmed as yet.
Host	Rabbit
Clonality	Polyclonal
Purity	Immunogen affinity purified serum in PBS pH 7.4.
Format	Lyophilized
Quantity	50 μg
Reconstitution	For reconstitution add 50 µl of sterile water
Storage	Store lyophilized/reconstituted at -20 °C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution	1 : 5000-10 000 (WB)
Expected apparent MW	23 kDa in legumes, 24 kDa (Arabidopsis thaliana)
Confirmed reactivity	Arabidopsis thaliana, Brassica oleracea, Hordeum vulgare, Medicago truncatula, Pisum sativum, Spinacia oleracea
Predicted reactivity	Species of your interest not listed? Contact us
Additional information	Note, the calculated molecular weight of pea ferritin is 28 kDa. Removal of the N-terminal targeting sequence upon protein import into plastids results in a protein with an apparent mol weight of ~23 kDa.
	This antibody is also recognizing horse ferritin (above 100 ng in Western blot).
Selected references	<u>Jiang</u> et al. (2022) Reactive effects of pre-sowing magnetic field exposure on morphological characteristics and antioxidant ability of Brassica juncea in phytoextraction. Chemosphere. 2022 Sep;303(Pt 1):135046. doi: 10.1016/j.chemosphere.2022.135046. Epub 2022 May 23. PMID: 35618056. <u>Bastow</u> et al. (2018). Vacuolar Iron Stores Gated by NRAMP3 and NRAMP4 Are the Primary Source of Iron in Germinating Seeds. Plant Physiol. 2018 Jul;177(3):1267-1276. doi: 10.1104/pp.18.00478.

Application example



Molecular weight markers (1); purified ferritin from *Pisum sativum*, 5 ng (2) and 0.5 ng (3); 5 μ g of Pisum sativum total cell extract (4); 5 μ g *Arabidopsis thaliana* total leaf extract (5). Proteins were separated on a 12% SDS-PAGE gel and transferred to nitrocellulose membrane using a semi-dry blotting apparatus. Blots were blocked in TBS, 0.1% (v/v) Tween-20, 5% (w/v) skimmed dried milk (TBS-TM) for 1 hour at RT. The antiserum was diluted 1: 5,000 in TBS-TM and incubated with the blot for 2 hours at RT. The blot was washed 3 times for 10 min with TBS-TM, then incubated with secondary antibodies anti-rabbit IgG HRP diluted to 1: 5000 in TBS-T for 1 hour. The blot was washed 4 times with TBS-T and developed with chemiluminescent detection reagent.

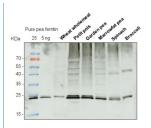
Courtesy of Dr. Janneke Balk, John Innes Centre, UK



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Molecular weight markers, purified pea ferritin and 20 μ g of plant cell extracts from various plant species depicted at the top of the image. Proteins were separated on a 12% SDS-PAGE gel and transferred to nitrocellulose membrane using a semi-dry blotting apparatus. Blots were blocked in TBS, 0.1% (v/v) Tween-20, 5% (w/v) skimmed dried milk (TBS-TM) for 1 hour at RT. The antiserum was diluted 1: 5,000 in TBS-TM and incubated with the blot for 2 hours at RT. The blot was washed 3 times for 10 min with TBS-TM, then incubated with secondary antibodies anti-rabbit IgG HRP diluted to 1: 5000 in TBS-T for 1 hour. The blot was washed 4 times with TBS-T and developed with chemiluminescent detection reagent.

Courtesy of Emily Jones, John Innes Centre, UK