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Product no AS07 217

FtsZ | Procaryotic cell division GTPase (cyanobacterial)

Product information

Immunogen	Whole cyanobacterial (*Anabaena* PCC 7120) FtsZ protein, UniProt: Q3MC27 overexpressed in *E.coli*.
Host	Rabbit
Clonality	Polyclonal
Purity	Serum
Format	Lyophilized
Quantity	50 µl
Reconstitution	For reconstitution add 50 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
Additional information	To detect *E.coli* FtsZ protein we recommend a following product: AS10 715 | anti-FtsZ procaryotic cell division GTPase (bacterial), rabbit antibody **To detect FtsZ protein in higher plants** following antibodies are recommended: AS09 413 | Anti-FtsZ1 and 2 | Plant cell division protein FtsZ1 and FtsZ2, rabbit antibodies AS13 2651 | Anti-FtsZ2 | Plant cell division protein ftsZ2, rabbit antibodies

Application information

Recommended dilution	1 : 200 (IL) Immunogold-TEM, 1 : 500 (IF), 1 : 2000-1 : 5000 (WB)
Expected | apparent MW	44.5 | 50 kDa
Confirmed reactivity	*Cylindrospermopsis raciborskii CS-505, Listeria monocytogenes (weak reaction), Synechococcus elongatus*
Predicted reactivity	*Phaeodactylum tricornutum*, *Prochlorococcus* sp.
Not reactive in	higher plants
Additional information	This antibody can be used as a loading control antibody in cyanobacteria. Immunofluorescence has been done by labelling *Synechococcus elongatus* cells at 30°C for 2 hours with FtsZ antibodies diluted to 1: 500 in blocking buffer. Detection images can be found in Kabeya et al (2010). This product can be sold containing ProClin if requested.
Selected references	Kurmayer et al. (2020). Chemically labeled toxins or bioactive peptides show a heterogeneous intracellular distribution and low spatial overlap with autofluorescence in bloom-forming cyanobacteria. Sci Rep. 2020 Feb 17;10(1):2781. doi: 10.1038/s41598-020-59381-w. (Immunofluorescence) Zhan et al. (2018). Photobleaching Enables Super-resolution Imaging of the FtsZ Ring in the Cyanobacterium Prochlorococcus. J Vis Exp. 2018 Nov 6;(141). doi: 10.3791/58603. MacCready et al. (2016). Robust Min-System Oscillation in the Presence of Internal Photosynthetic Membranes in Cyanobacteria. Molecular Microbiology November 5 2016. doi: 10.1111/mmi.13571 Probst et al. (2014). Biology of a widespread uncultivated archaeon that contributes to carbon fixation in the subsurface. Nat Commun. 2014 Nov 26;5:5497. doi: 10.1038/ncomms6497. Miyagishima et al. (2014). DipM is required for peptidoglycan hydrolysis during chloroplast division. BMC Plant Biol. 2014 Mar 6;14(1):57. (immunofluorescence) Plominsky et al. (2013). Dinitrogen Fixation Is Restricted to the Terminal Heterocysts in the Invasive Cyanobacterium Cylindrospermopsis raciborskii CS-505. PLOS ONE, Open Access. Kabeya et al (2010). The YlmG protein has a conserved function related to the distribution of nucleoids in chloroplasts and cyanobacteria. BMC Plant Biology 10:57.

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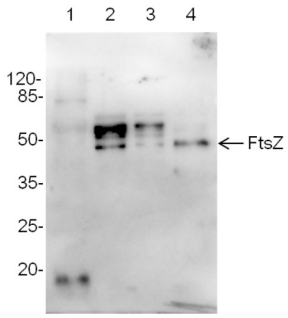
Application example



Total protein samples (5 or 10 µg) from: *Arabidopsis thaliana,* leaf **(1)**, *Synechocystis* 6803 motile **(2)**, *Synechocystis* 6803 GT (glucose tolerant strain) **(3)**, *Synechococcus elongates* 7942 **(4)**, Marker - Pierce™ Prestained Protein MW Marker (kat #26612) were extracted with buffer (10mM Tris HCl, pH 8.0, 0.5% LDS, 4% glycerol, 0.1 mM EDTA) were mixed with sample buffer and denatured for 5 min at 95°C. Samples were separated on 10% SDS -PAGE and blotted 1h to nitrocellulose membrane (Amersham Protran) using sem i-dry transfer (Bio -Rad) in standard transfer buffer in presence of 10% methanol. Transfer of proteins to the membrane was checked using 0,5% Ponceau S staining before the blocking step. Blots were blocked in buffer (2% low -fat milk in 1xPBS, 0,1% Tween) for 1h at room temperature (RT) with agitation. Blots were incubated in the primary antibody at a dilution of 1: 1000 at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in PBS -T at RT with agitation. Blot was incubated in secondary antibody (goat anti -rabbit IgG, AS09 602, Agrisera) diluted to 1:30 000 for 1h at RT with agitation. The blot was washed as above and developed for 5 min with chemiluminescence detection reagent and ChemiDoc detection system.

Courtesy Dr. Elena Pojidaeva, Laboratory of Plant Gene Expression, Timiryazev Institute of Plant Physiology RAS, 127276 Moscow Russia