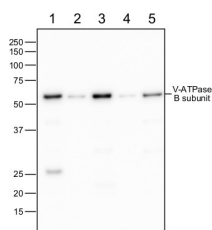


Product no **AS14 2775****V-ATPase, B | vacuolar H<sup>+</sup>-ATPase subunit B****Product information**

<b>Immunogen</b>	KLH-conjugated synthetic peptide chosen from <i>Arabidopsis thaliana</i> V-ATPase subunit B, isoform B1: UniProt: <a href="#">Q683E8</a> , TAIR: <a href="#">AT1G76030</a> , isoform B2 UniProt: <a href="#">Q9SZN1</a> , TAIR: <a href="#">AT4G38510</a> , isoform B3: UniProt: <a href="#">Q8W4E2</a> , TAIR: <a href="#">AT1G20260</a>
<b>Host</b>	Rabbit
<b>Clonality</b>	Polyclonal
<b>Purity</b>	Immunogen affinity purified serum in PBS pH 7.4.
<b>Format</b>	Lyophilized
<b>Quantity</b>	50 µg
<b>Reconstitution</b>	For reconstitution add 50 µl of sterile water
<b>Storage</b>	Store lyophilized/reconstituted at -20°C; make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
<b>Additional information</b>	Antibodies will detect target protein in a few µg of a crude preparation loaded per well. If purified preparations of vacuolar and plasma membranes are used, one µg load per well should be sufficient

**Application information**

<b>Recommended dilution</b>	1 : 1000 (WB)
<b>Expected   apparent MW</b>	53   57 kDa ( <i>Vigna radiata</i> )
<b>Confirmed reactivity</b>	<i>Arabidopsis halleri</i> , <i>Nicotiana tabaccum</i> , <i>Thellungiella salsuginea</i> , <i>Vigna radiata</i>
<b>Predicted reactivity</b>	<i>Acetabularia sp.</i> , <i>Arundo donax</i> , <i>Chlamydomonas reinhardtii</i> , <i>Cucumis sativus</i> , <i>Glycine soja</i> , <i>Gossypium mexicanum</i> , <i>Halostachys caspica</i> , <i>Haloxylon ammodendron</i> , <i>Hordeum vulgare</i> , <i>Medicago truncatula</i> , <i>Mesembryanthemum crystallinum</i> , <i>Ostreococcus tauri</i> , <i>Oryza sativa</i> , <i>Panax ginseng</i> , <i>Physcomitrium patens</i> , <i>Pinus sylvestris</i> , <i>Populus trichocarpa</i> , <i>Pyrus sp.</i> , <i>Ricinus communis</i> , <i>Theobroma cacao</i> , <i>Triticum aestivum</i> , <i>Zea mays</i> , <i>Zostera marina</i>
	Species of your interest not listed? <a href="#">Contact us</a>
<b>Not reactive in</b>	<i>Thermatoga neapolitana</i>
<b>Additional information</b>	Protein or membrane sample should be treated at 70°C for 10 min before loading on the gel

**Applicaition example**

Proteins were separated by SDS-PAGE and transferred to an Immobilon-P membrane (Millipore) using Trans-Blot SD Semi-Dry Transfer Cell (Bio-Rad) with transfer buffer (100 mM Tris, 192 mM Glycine, 0.02% (w/v) SDS and 5% (v/v) methanol). After treatment with 1% blocking agent, the membrane filter was incubated with the primary antibody (1:1000) and then with horseradish peroxidase (HRP)-conjugated goat anti-rabbit IgG (H+L) (Agrisera [AS09 602](#), 1:25 000). Chemiluminescent reagent was used for detection of antigens. Chemiluminescence was detected with a Light-Capture II imaging device with a cooled CCD camera (Atto).

**Samples:**

- 1: 10 µg of 100,000 x g precipitate prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 2: 0.2 µg of vacuolar membrane enriched fraction prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 3: 2 µg of vacuolar membrane enriched fraction prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 4: 0.2 µg of vacuolar membrane enriched fraction prepared from *Vigna radiata* 4 days old hypocotyls. 5: 2 µg of vacuolar membrane enriched fraction prepared from *Vigna radiata* 4 days old hypocotyls.

# Agrisera

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contact: [support@agrisera.com](mailto:support@agrisera.com)

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | [www.agrisera.com](http://www.agrisera.com)

fraction prepared from *Vigna radiata* 4 days old hypocotyls.

Courtesy of Drs. Masayoshi Maeshima and Dr Shoji Segami, Nagoya University, Japan