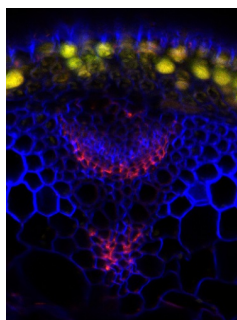


Product no **AS16 3136-1ml****Fucosylated xyloglucan (clone CCRC-M1)****Product information**

<b>Immunogen</b>	MeBSA-conjugated sycamore rhamnogalacturonan (non-covalent)
<b>Host</b>	Mouse
<b>Clonality</b>	Monoclonal
<b>Subclass/isotype</b>	IgG1
<b>Purity</b>	Cell culture supernatant.
<b>Format</b>	Liquid
<b>Quantity</b>	1 ml
<b>Storage</b>	Antibody can be stored up to 1 month at 4°C, and at -80°C for up to 1 year. Make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
<b>Additional information</b>	Exact working dilution needs to be determined by end user, Epitope structure for carbohydrate antigen is: alpha-Fuc-(1,2)-beta-Gal

**Application information**

<b>Recommended dilution</b>	1 : 10 or nondiluted (ELISA, (IF), (IHC)
<b>Confirmed reactivity</b>	<i>Acer pseudoplatanus</i> , <i>Arabidopsis thaliana</i>
<b>Predicted reactivity</b>	Dicots Species of your interest not listed? <a href="#">Contact us</a>
<b>Not reactive in</b>	No confirmed exceptions from predicted reactivity are currently known
<b>Selected references</b>	<a href="#">Pattathil</a> et al. (2012). Immunological approaches to plant cell wall and biomass characterization: Glycome Profiling. <i>Methods Mol Biol.</i> 2012;908:61-72.doi: 0.1007/978-1-61779-956-3_6. <a href="#">Patathil</a> et al. (2010). A comprehensive toolkit of plant cell wall glycan-directed monoclonal antibodies. <i>Plant Physiol.</i> 2010 Jun;153(2):514-25.doi: 10.1104/pp.109.151985.

**Application example**

Localization of fucosylated xyloglucan (red) in *Arabidopsis thaliana* hypocotyl, Calcufluor White counterstain (blue) and cell wall autofluorescence (yellow).

The 31 days-old hypocotyls were immersed in 150  $\mu$ L PME fixation buffer (25 mM PIPES, 1 mM  $MgSO_4$ , 1 mM EGTA) and then subjected to three consecutive cycles of 5 min-long vacuum infiltration (21°C, 68 kPa). Afterwards they were washed three times in PME (21°C, 68 kPa) prior to storage at 4°C in PME. Hypocotyls were encased in 1 cm<sup>3</sup> blocks of 5% agar at 65°C, and stored at 4°C to set. Transverse 40  $\mu$ m thick sections were cut from segments using a VT100S vibrating microtome (Leica) and blocked for at least 1 h in 5% bovine serum albumin in TBST. Blocking solution was discarded and sections were incubated at 4°C for 16 h with 5  $\mu$ l of the anti-Fucosylated xyloglucan antibody, followed by 2 washes in 100  $\mu$ L TBST. Sections were then incubated for 1 h at 21°C in the dark in 10  $\mu$ l of 2  $\mu$ g/ $\mu$ l Alexa Fluor<sup>TM</sup> 568 donkey anti-mouse IgG (H+L; 1:36). Sections were again washed twice in 40  $\mu$ L TBST prior to counter-staining with 0.015% Calcufluor White (Sigma-Aldrich). Sections were again washed twice in 100  $\mu$ L TBST to remove excess counter-stain and unbound secondary antibody. Immunofluorescence of AlexaFluor 568 was excited with a 561 nm laser, and emitted light filtered at 575–600 nm, while Calcufluor White was subsequently scanned on an independent channel with a 405 nm laser and emission observed at 420–430 nm using laser scanning microscope Zeiss LSM780 point-scan system at 1024  $\times$  1024 pixels (pixel size, 0.6–0.83  $\mu$ m) with a 10X objective.

Courtesy Dr. Urs Fisher, Umeå Plant Science Centre, Sweden